

Microbial Air Sampling Plans in Cleanrooms According to Regulatory Authorities

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ABSTRACT

The reported specific “Sampling Plan” has been prepared according to the requests of the Annex 1–Manufacture of Sterile Medicinal Products – Environmental Monitoring – Viable Monitoring. The Chapter 9.27 says that: “Continuous monitoring in grade A and B areas should be undertaken for the full duration of critical processing, including equipment (aseptic set up) assembly and filling operations (i.e.: an understanding of functions and interactions of each clean area). The monitoring should be performed in such a way that all interventions, transient events and any system deterioration would be captured, and any risk caused by interventions of the monitoring operations is avoided”. The document reports some examples of applications of the continuous monitoring of Cleanroom for viable microorganisms.

Keywords: Air Sampler; Aspirating Chamber; Aspirating Head; At Rest; Cleanroom; Cfu, Contamination; Bacteria; Culture Plate; Delay; Disinfection; End Of Shift Activity; Fraction Time; Fraction Number; Incubation; In Operation; Microorganisms; Petri Dish; Run, Standard; Sterilization; Unidirectional Laminar Flow

Introduction

USP guidelines for Cleanrooms state that the contamination monitoring is performed during the normal working day activity of each shift. The volume of 1000 litres of air should be collected at each sampling point. Air samples should be collected “at rest”, “in operation”, and “after process”.

Sampling Plan

The sampling plan, part of a SOP (Standard Operating Procedures), should be written prior to begin an ongoing environmental program. A robust air sampling program is and developed based upon the size of the area, number of hoods and after a risk assessment. In order to trend the number and type of microorganisms on a day / weekly / monthly basis, it is necessary to sample with the same method / protocol each time. Areas to be sampled are indicated on a floor plan of the Cleanroom and should include laminar flow benches, BSC, Isolator, RABS, buffer zone, gowning areas, and anteroom areas.

1000 L Of Air to Be Collected

The operators, employing a process involving a single head active air sampler, will need to perform several manipulations

regarding preparation and culture plate transfer. Working time and the risk of contamination can be reduced with the use of a two or three aspirating head format sampler. The sampler can be prepared and programmed prior to starting daily activity and the culture plates collected at the end of the shift. “Shift working time” differs from company to company, and with cyclical processing circumstances, from time to time. In this document, we report several examples using TRIO.BAS DUO and TRIO.BAS TRIO to demonstrate sampling flexibility. In addition to the programming samples provided, note instrument start times may also be programmed with the “delay” function (Figures 1 & 2).



Figure 1.



Figure 2.

Materials

Microbial air sampler with 2 aspirating heads, 100mm Petri dish with TSA agar culture media, “Daily Shift” sterile certified aspirating heads.

Protocol

Hours long sampling spans can be programmed by different “delay”, “fraction time”, and “fraction number”. Are here reported several examples: 1.000 litres of air samples during different working shift runs. The cycles are stored in the TRIO.BAS air sampler memory and then can be transferred to a P.C., via one of the data transfer accessory options (Tables 1 & 2).

Table 1.

EXAMPLE OF 2h.20min WORK SHIFT						
SAMPLER CFG: VOL - 1000 LTS - OK - DELAY - 0 - OK - FRACTION NUM. - 2 - OK - FR. TIME - 60 - OK - HEAD MODE SQS - OK - GO - HEAD 1&2 - OK						
Duo 100 l/m prova 2 fraction - 1000 l/m						
Head	Fraction	Starting Hour	Time Test	Sampler %	L/m session	End of test
1	A	9	5,00 MIN	50%	500	9.05
1	B	10.05	5,00 MIN	100%	500	10.10
2	A	10.10	5,00 MIN	50%	500	10.15
2	B	11.15	5,00 MIN	100%	500	11.20

Table 2.

EXAMPLE OF 3h.21min WORK SHIFT						
SAMPLER CFG: VOL - 1000 LTS - OK - DELAY - 0 - OK - FRACTION NUM. - 3 - OK - FR. TIME - 45 - OK - HEAD MODE SQS - OK - GO - HEAD 1&2 - OK						
Duo 100 l/m prova 3 fraction - 1000 l/m						
Head	Fraction	Starting Hour	Time Test	Sampler %	L/m session	End of test
1	A	9.00	3,30 MIN	33%	333	9.03
1	B	9.48	3,30 MIN	66%	333	9.52
1	C	10.37	3,30 MIN	100%	333	10.40
2	A	10.40	3,30 MIN	33%	333	10.44
2	B	11.29	3,30 MIN	66%	333	11.32
2	C	12.17	3,30 MIN	100%	333	12.21

Standard Operating Procedure (Sop)

The SOP-Standard Operating Procedures for environmental monitoring can be downloaded or requested from ORUM International S.r.l.

Practical Application

Starting “At rest” at 09:00 / “In operation” at 10:05 / “At end” at 11:20a.m.

Starting “At rest” at 09:00a.m. “In operation” at 10:37 / “At end” at 12:21p.m.

The continuous monitoring of the viable microorganisms can be expanded up to 6 or 8 hours with the appropriate use of

the “delay”, “fraction number” and “fraction time” functions of the microbial air samplers.

Conclusion

This protocol conforms what the Regulatory Authorities request: “9.27 Continuous monitoring in grade A an B areas should be undertaken for the full duration of critical processing, including equipment, assembling and filling operation”.

References

1. Annex 1, Manufacture of Sterile Medicinal Products – Environmental Monitoring – Viable Monitoring.

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